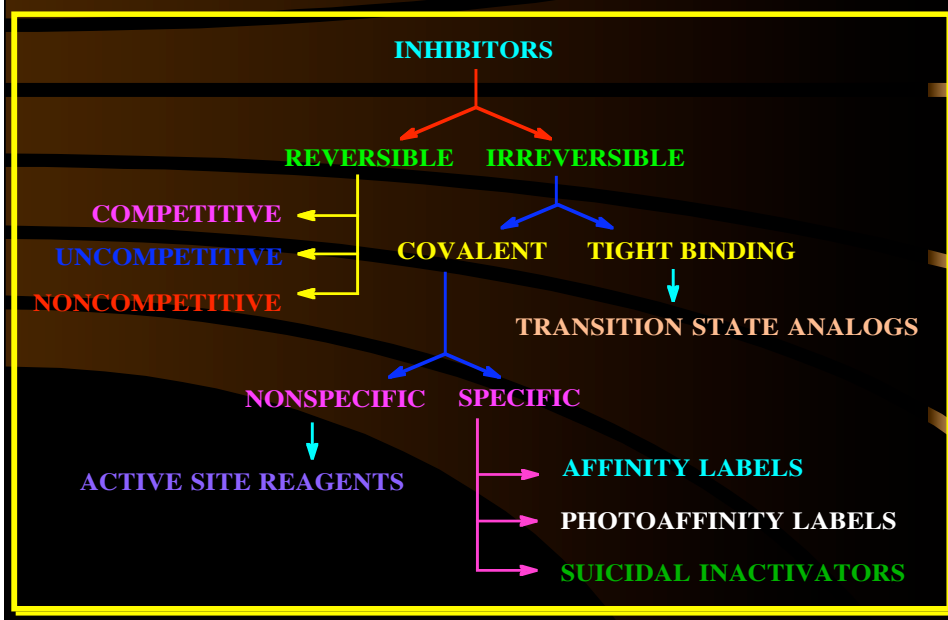


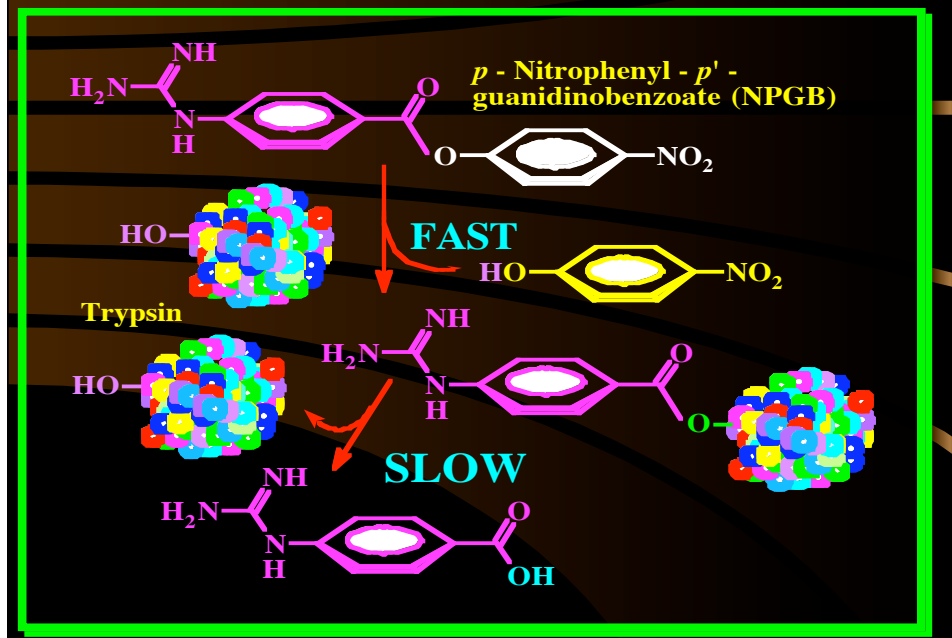
INHIBITORS

MANICKAM SUGUMARAN
PROFESSOR OF BIOLOGY
U. MASS - BOSTON
BOSTON, MA 02125

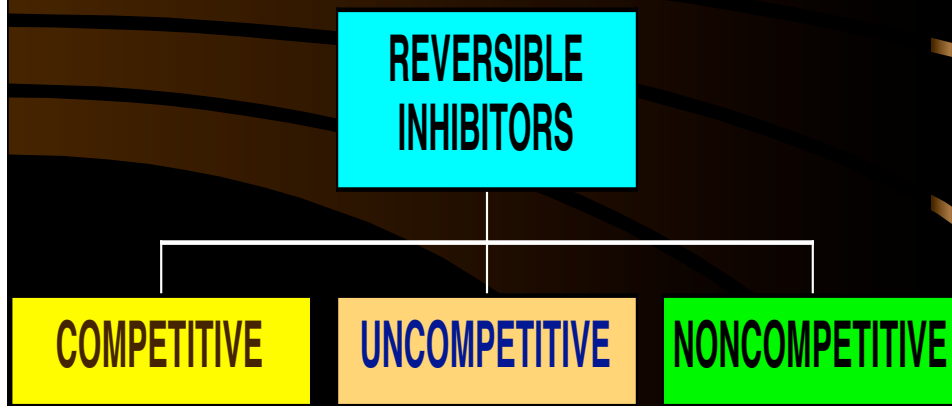
INHIBITORS - DIFFERENT TYPES

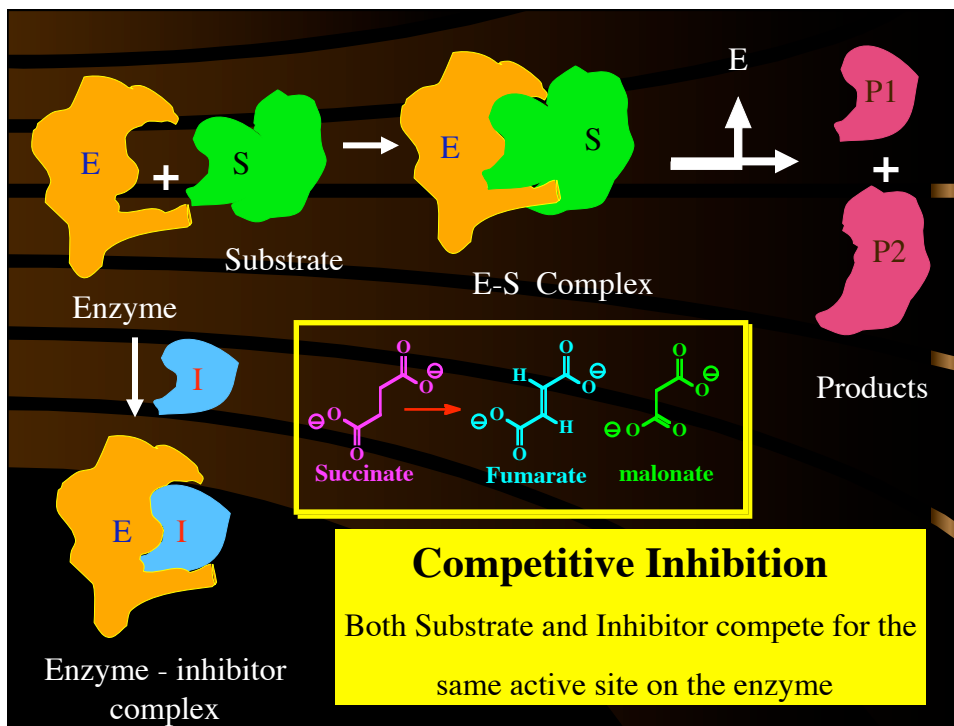


Active site titration of trypsin



THREE DIFFERENT REVERSIBLE INHIBITORS





Competitive Inhibitor

$$E + S \rightleftharpoons ES \longrightarrow E + P$$

$$\updownarrow I$$

$$EI$$

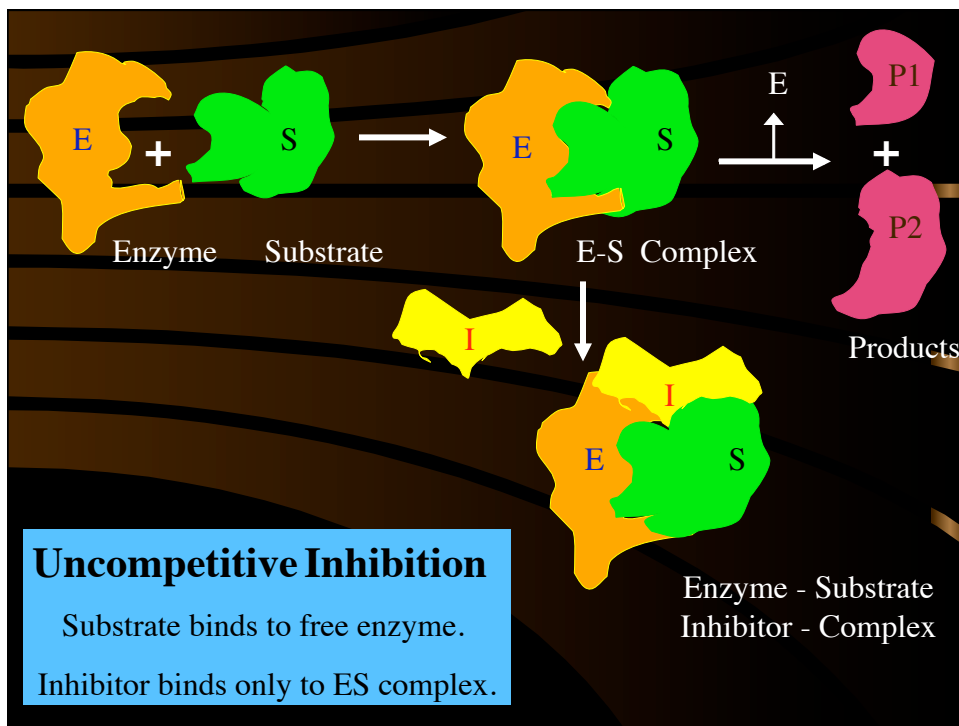
Both S and I compete for the same active site on the Enzyme

At very high [S] conc
There is no inhibition

Slope changes
Intercept on Y-axis
no change

Malonate is a competitive inhibitor of succinate dehydrogenase reaction

Competitive Inhibition



Uncompetitive Inhibitor

$$E + S \rightleftharpoons ES \xrightarrow{\quad} E + P$$

$$\quad \quad \quad \updownarrow I$$

$$\quad \quad \quad ESI$$

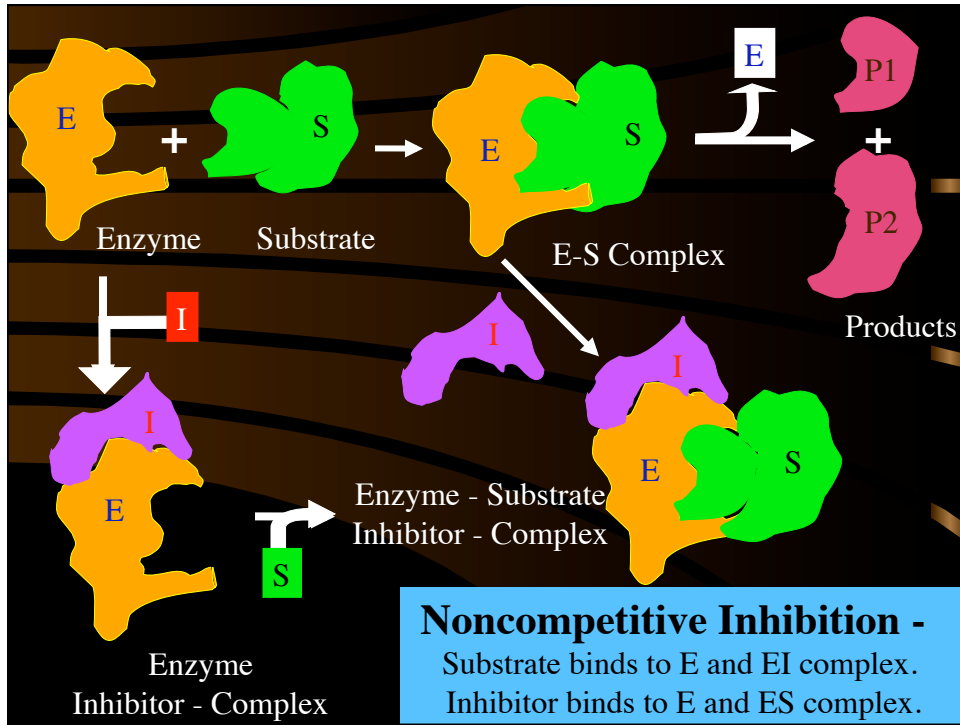
S reacts with E and I reacts
Only with ES complex

Slope: No change
Intercept on Y-axis:
Varies

Constant amount of ES is removed by I. Therefore, slope is constant

Uncompetitive Inhibition

The Lineweaver-Burk plot shows 1/v on the y-axis and 1/S on the x-axis. Four parallel lines are shown, representing different inhibitor concentrations: [I]^{'''} (red), [I]^{''} (green), [I]['] (blue), and No I (yellow). The lines are parallel, indicating uncompetitive inhibition, and their y-intercepts decrease as the inhibitor concentration increases.



Noncompetitive Inhibitor

$$\begin{array}{c}
 E + S \rightleftharpoons ES \longrightarrow E + P \\
 \updownarrow I \qquad \qquad \updownarrow I \\
 EI \xrightleftharpoons{S} ESI
 \end{array}$$

S reacts with E and EI complex.
I reacts with E and ES complex.

Noncompetitive Inhibition

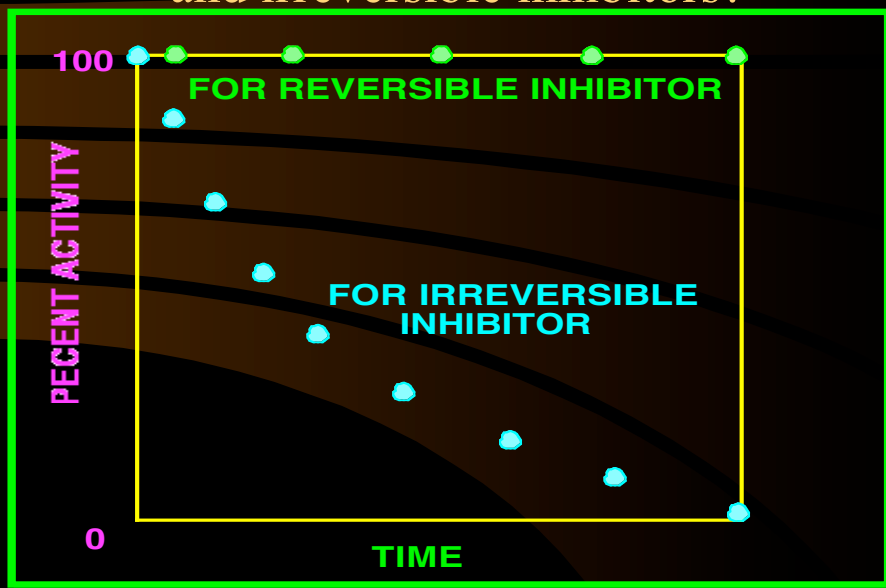
The Lineweaver-Burk plot shows the relationship between the reciprocal of the reaction velocity (1/v) on the y-axis and the reciprocal of the substrate concentration (1/S) on the x-axis. Four lines are shown, all intersecting at a single point on the negative x-axis. The lines represent different inhibitor concentrations: [I]''' (steepest), [I]'' (second steepest), [I]' (third steepest), and No I (shallowest). The y-intercept for all lines is positive, and it increases as the inhibitor concentration increases.

Both Slope and Intercept on Y-axis: Varies

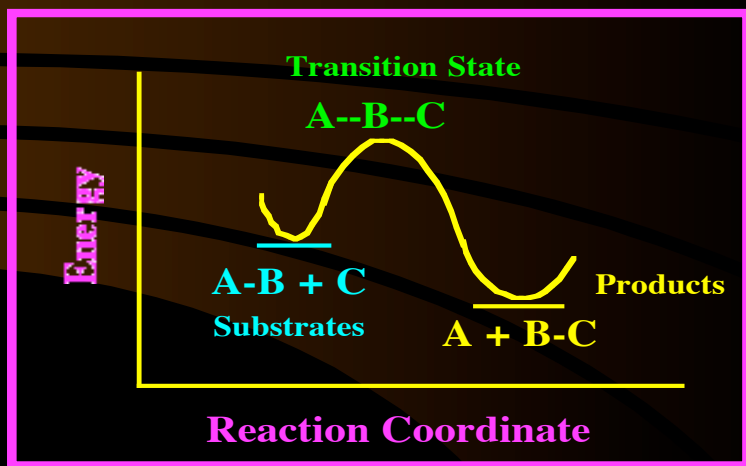
IRREVERSIBLE INHIBITORS ARE OF TWO TYPES



How to distinguish between reversible and irreversible inhibitors?



Tight Binding Inhibitors - Transition State Analogs

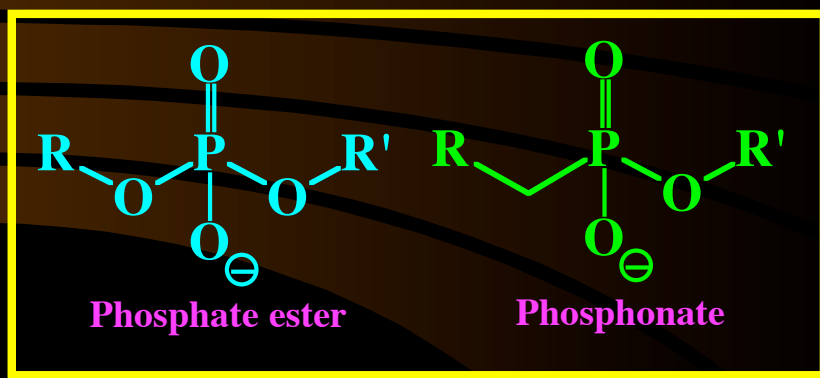


Transition State Analogs are Potent Enzyme Inhibitors

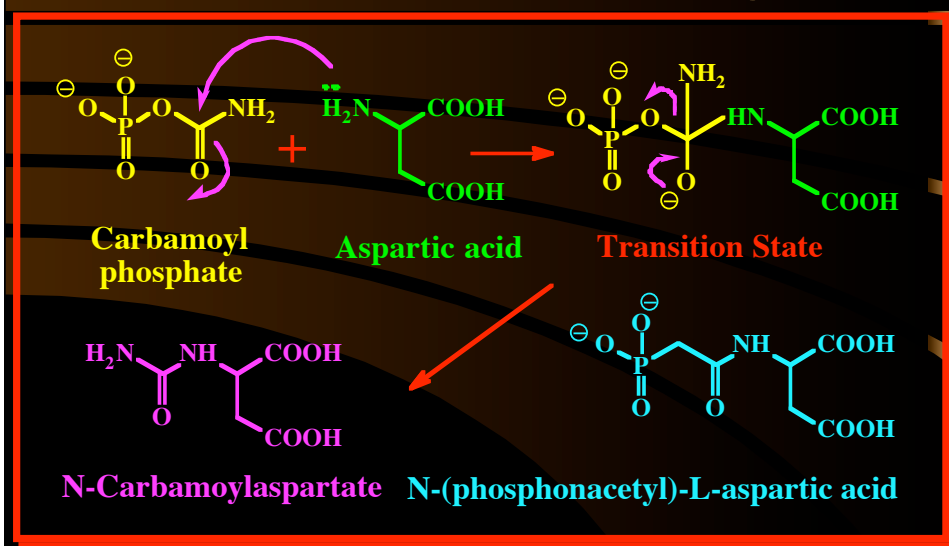
Compounds resembling the transition state
of enzyme catalyzed reactions
should be effective inhibitors of enzymes.

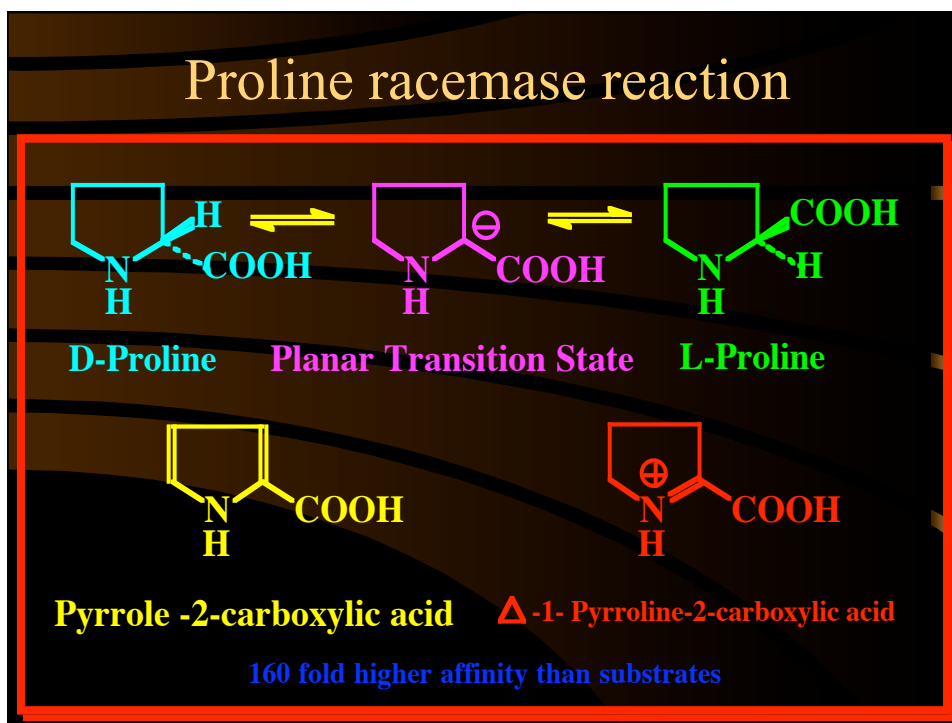
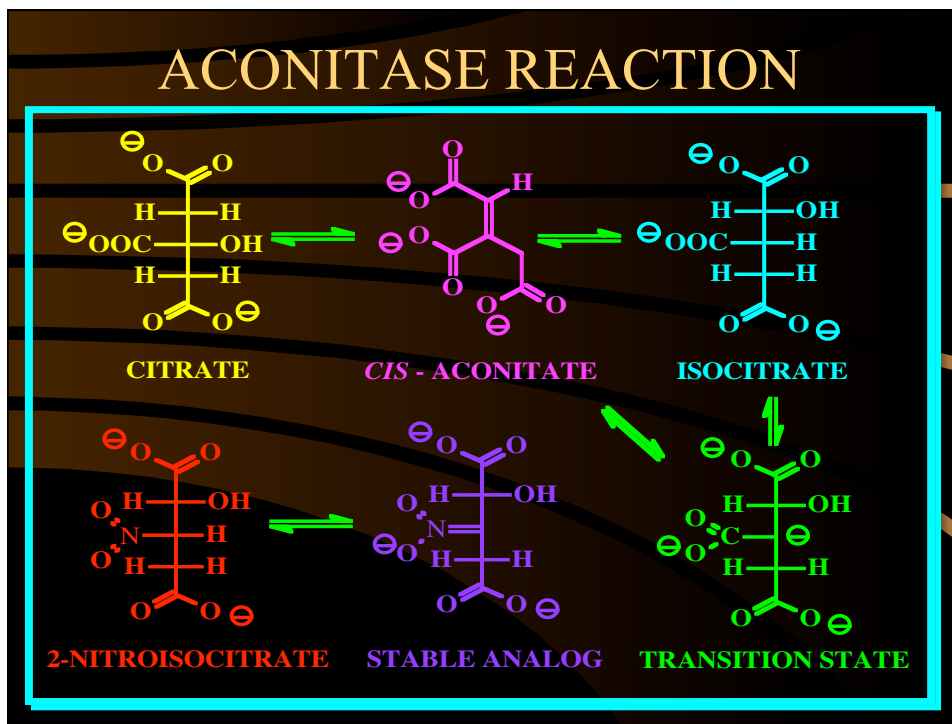
- Linus Pauling (1946)

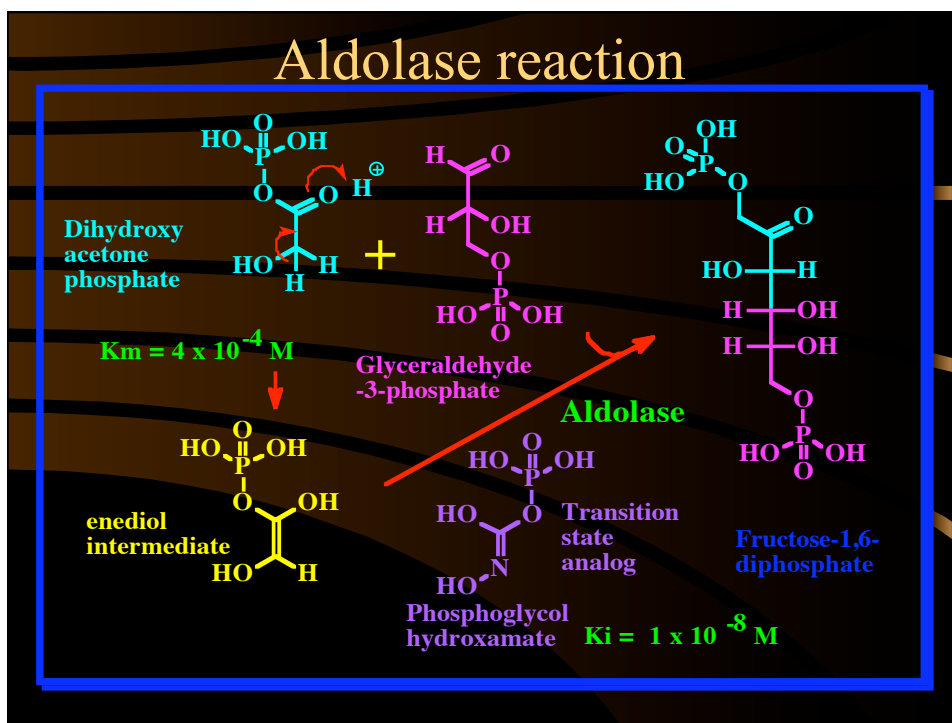
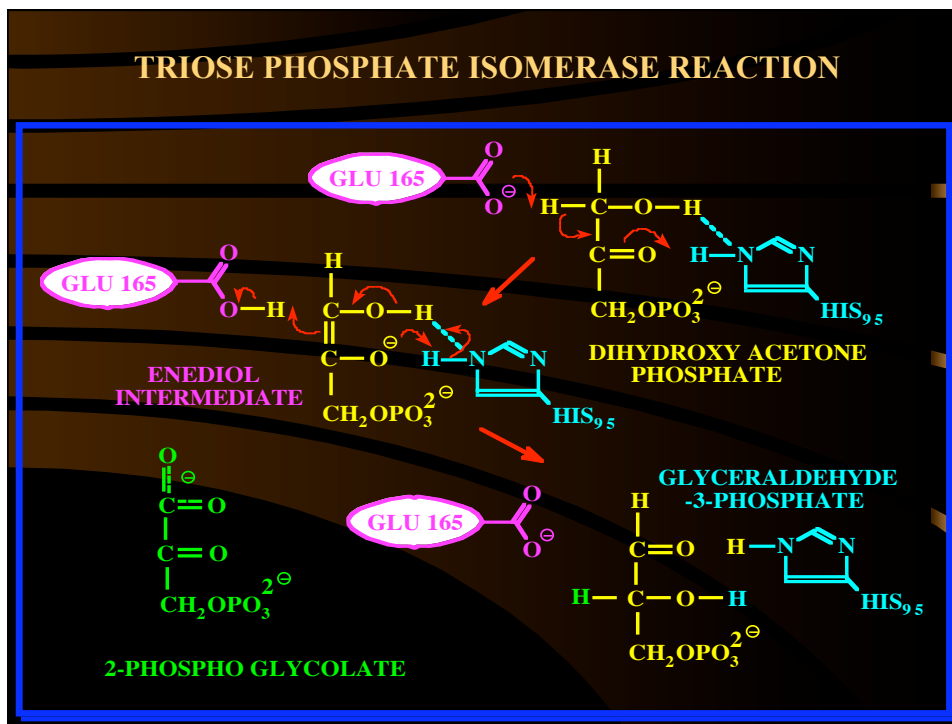
Phosphonates are good analogs
of phosphate esters



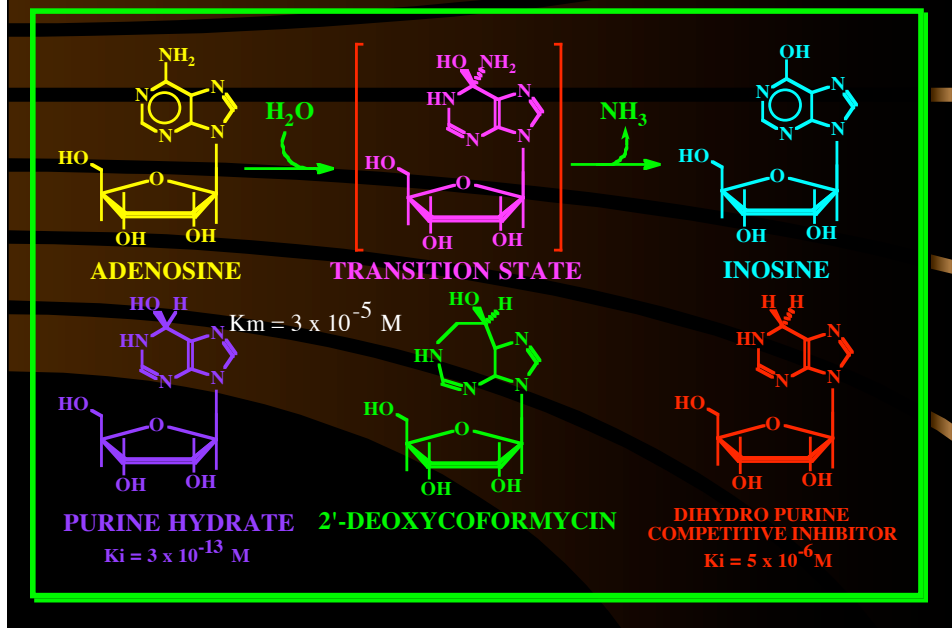
Aspartate transcarbamoylase and
its transition state analog



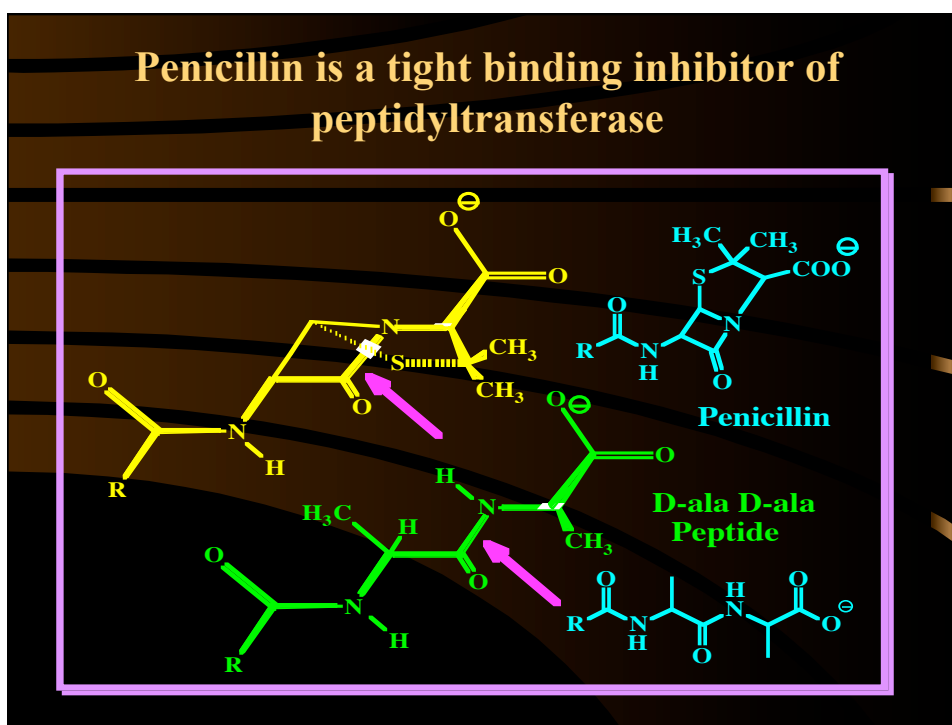




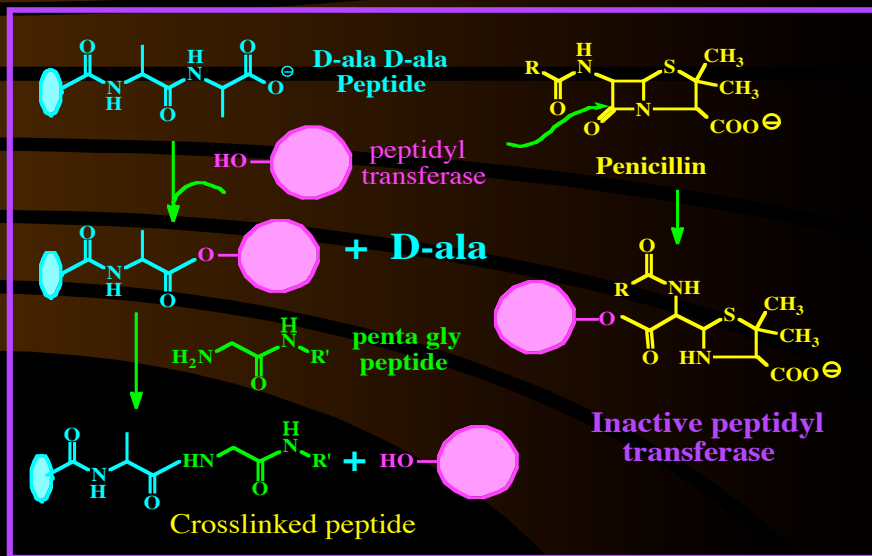
Transition state analog -Adenosine deaminase



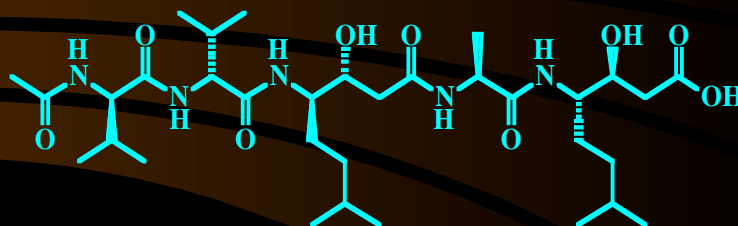
Penicillin is a tight binding inhibitor of peptidyltransferase



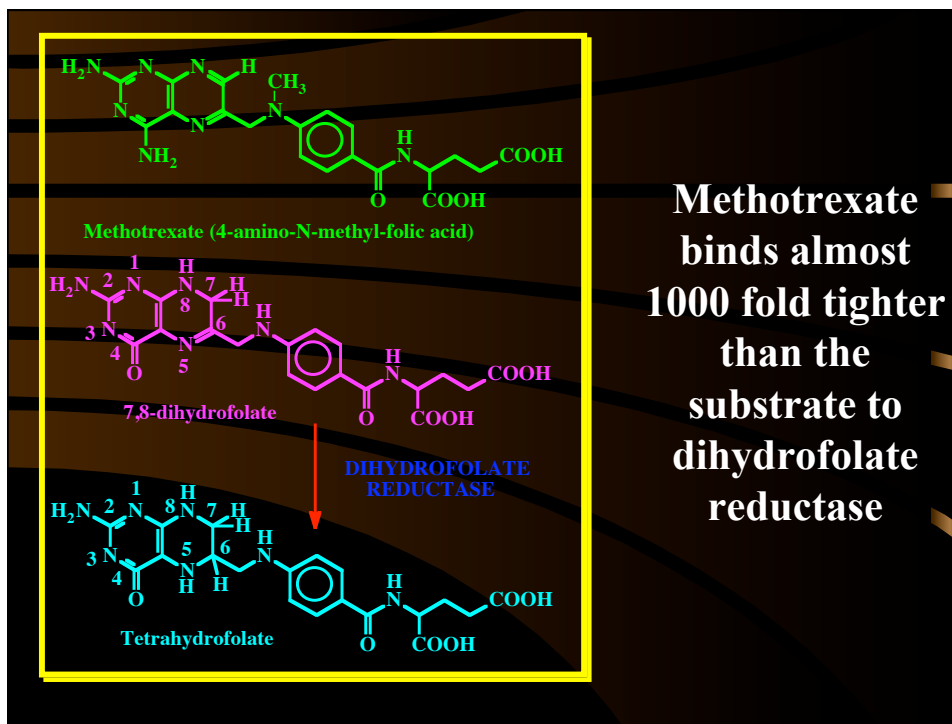
Penicillin inhibits transpeptidase reaction



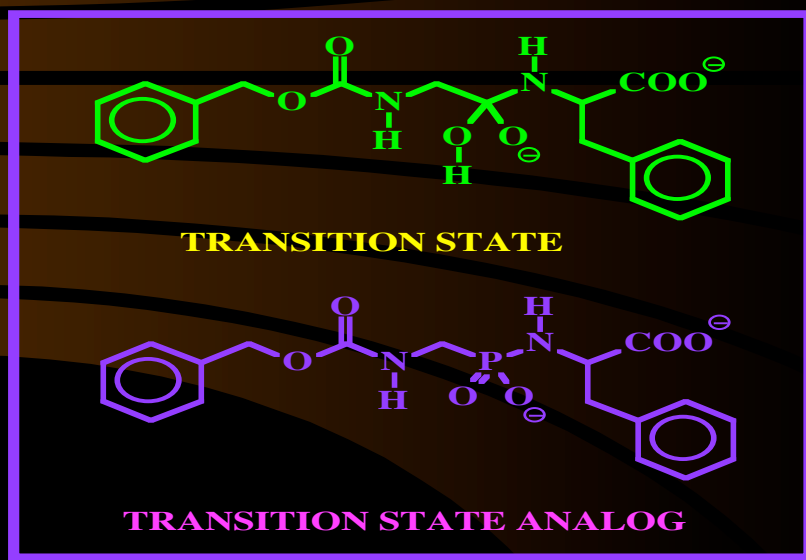
Pepstatin is a potent transition state analog of aspartyl proteases
(Hydroxyethylene group inside the peptide)



N-acetyl PEPSTATIN - A MICROBIAL PEPTIDE



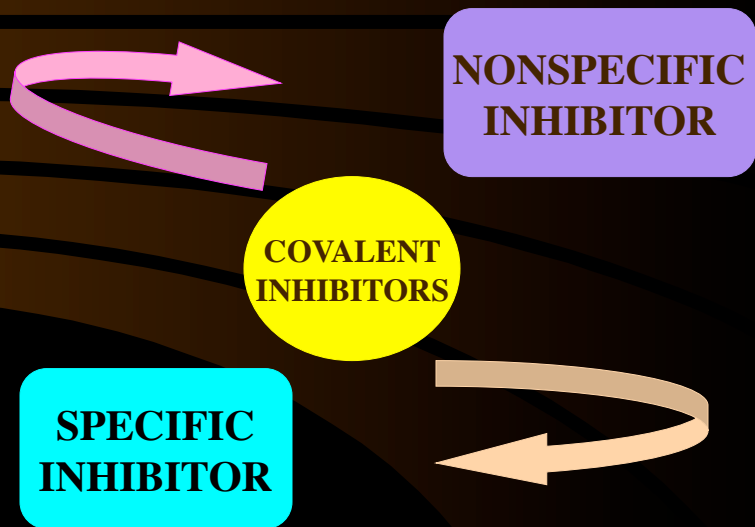
CARBOXYPEPTIDASE REACTION



OTHER TRANSITION STATE INHIBITORS

- **LYSOZYME - LACTONE**
- **RIBONUCLEASE - URIDINE VANADATE**
- **ISOCITRATE LYASE - 3-NITRO PROPIONATE**
- **RIBULOSE BISPHOSEPHATE CARBOXYLASE - CARBOXY ARABINITOL 1,5-BISPHOSEPHATE**
- **GLUTAMINE SYNTHETASE - METHIONINE SULFOXIMINE**
- **GAMMA -GLUTAMYL CYSTEINYL SYNTHETASE - S-(n-BUTYL)-HOMOCYSTEINE SULFOXIMINE**

THERE ARE TWO KINDS OF COVALENT INHIBITORS



Nonspecific Covalent Inhibitors

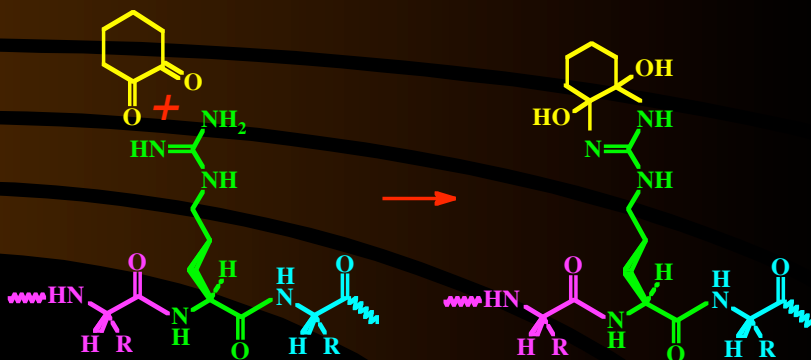
They react invariably with any enzyme as long as a particular reactive amino acid is present at or near the active site.
Generally the reaction leads to inactivation of the enzyme.

Best examples are active site reagents.

Serine - organophosphorus compounds
 Histidine, cysteine and Lysine - Haloacetates, Halomethylketones
 Histidine - diethylpyrocarbonate
 Arginine - 1,2-diketocyclohexane
 Cysteine - Thiol reagents, pCMB, DTNB
 Metal ion containing enzymes - cyanide, azide, carbon monoxide
 Carbonyl groups - reduction, hydroxylamine
 Lysine - anhydrides
 Carboxyl groups - esterification.

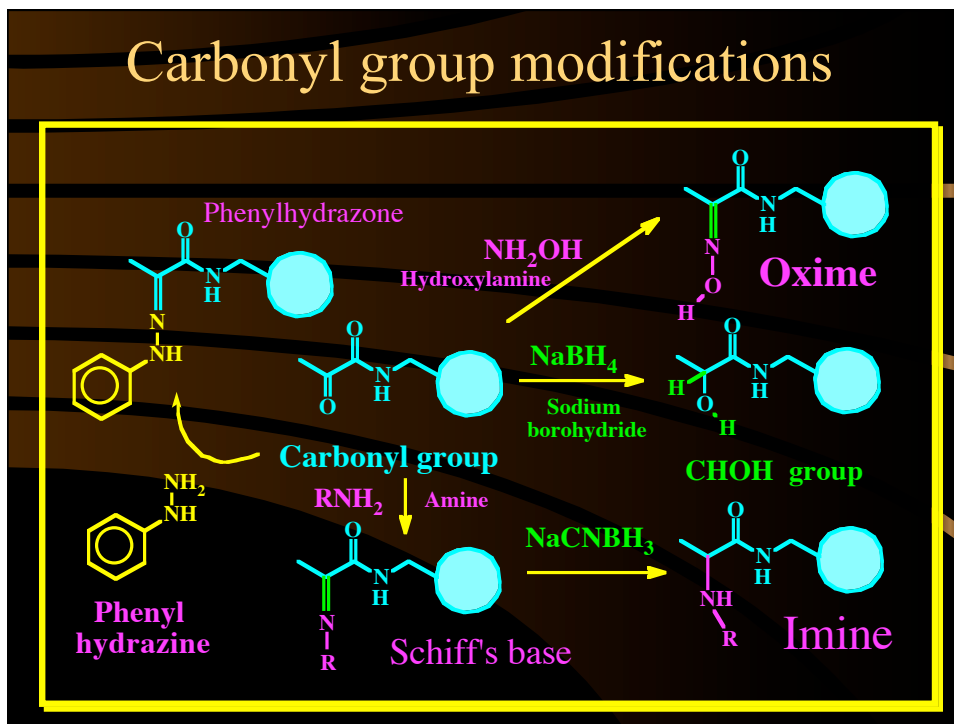
Modification of arginine

Reversible blocking of Arginine by cyclohexadione.
 E. L. Smith. *Methods in Enzymology* 47, 156-161 (1977).

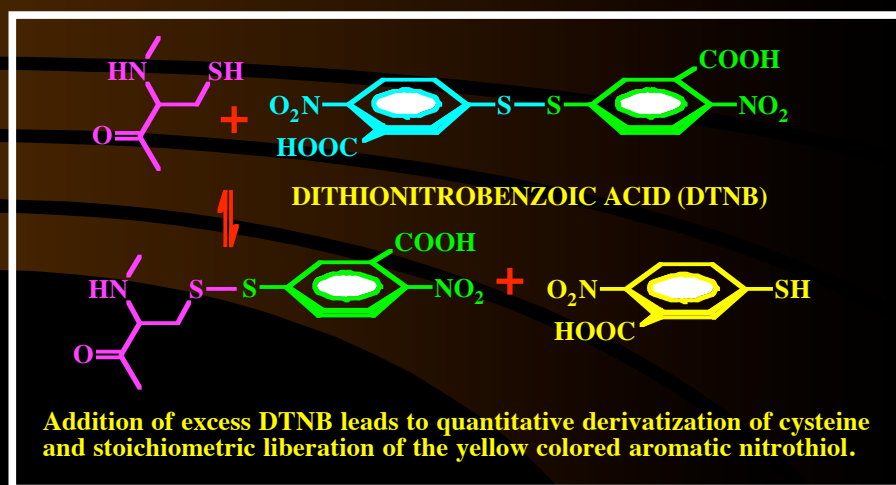


Arginine is modified by 1,2-cyclohexadione (or phenylglyoxal).
 The vicinyl dihydroxy adduct can be stabilized by borate.

Carbonyl group modifications

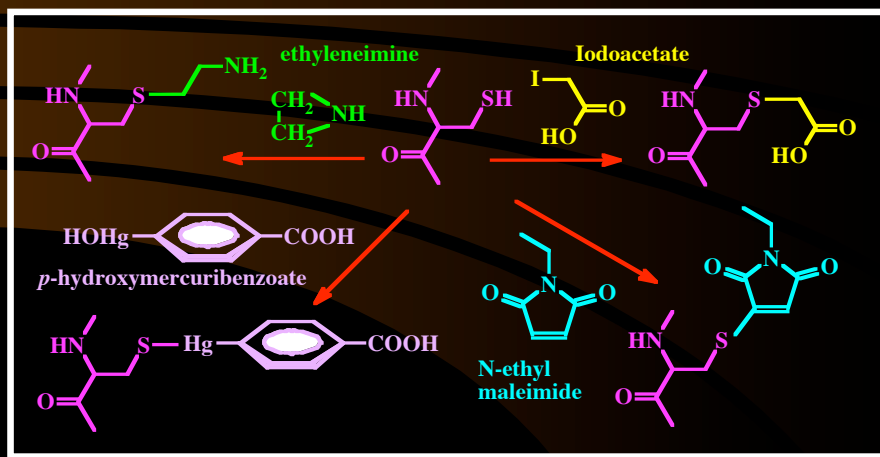


Cysteine modification (DTNB)

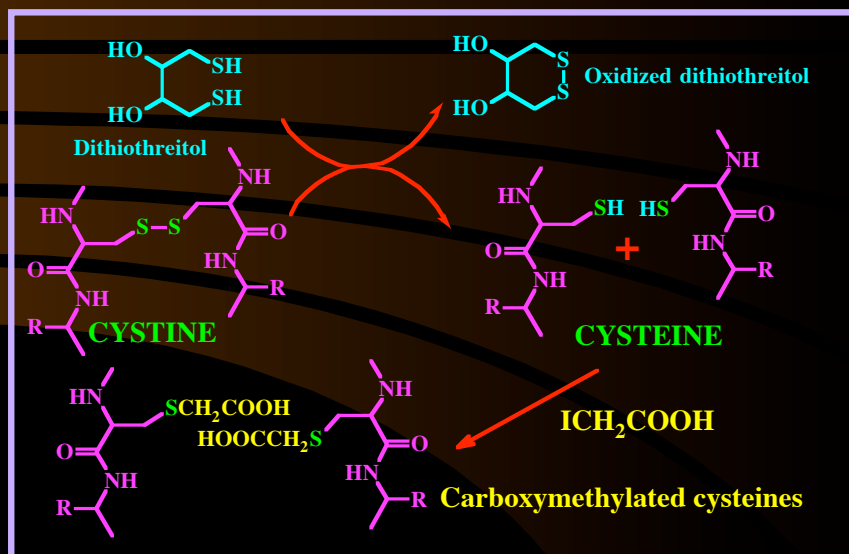


A number of reagents react with cysteine. Ethyleneimine, NEM, iodoacetate (as well as iodoacetamide) and *p*-hydroxymercuribenzoate

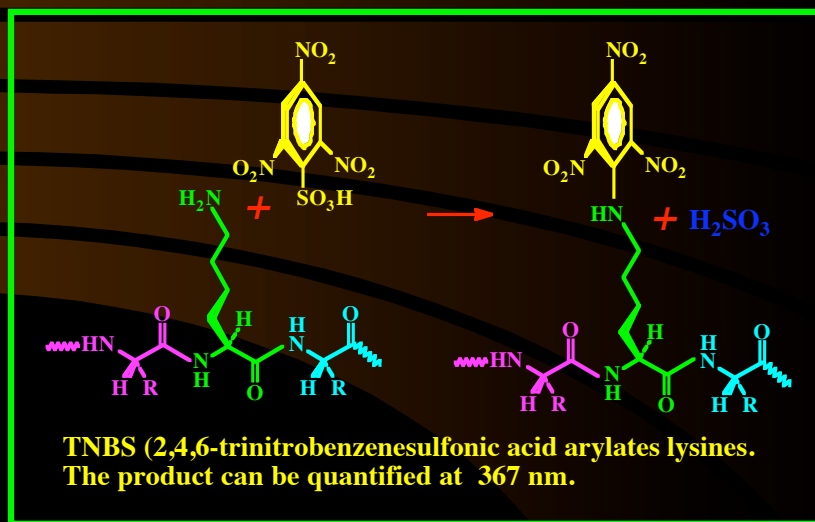
will modify cysteine as shown below .



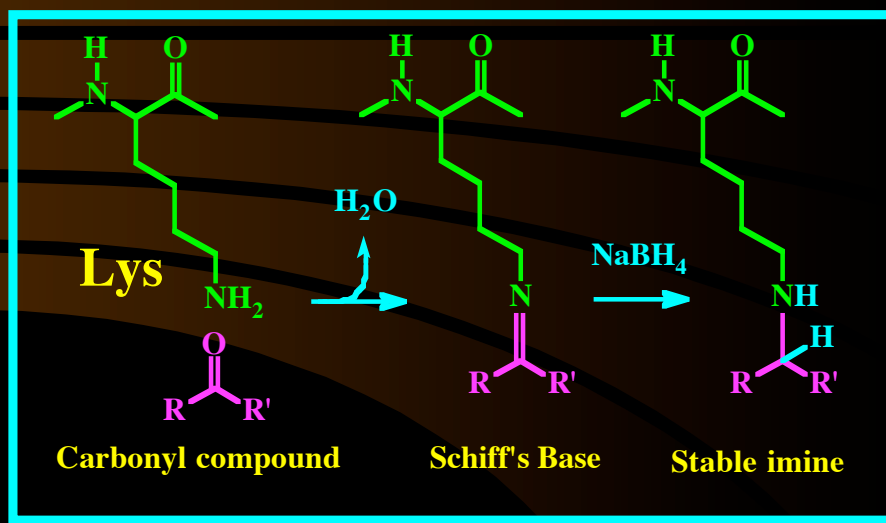
Dithiothreitol reduces disulfides and iodoacetate blocks them



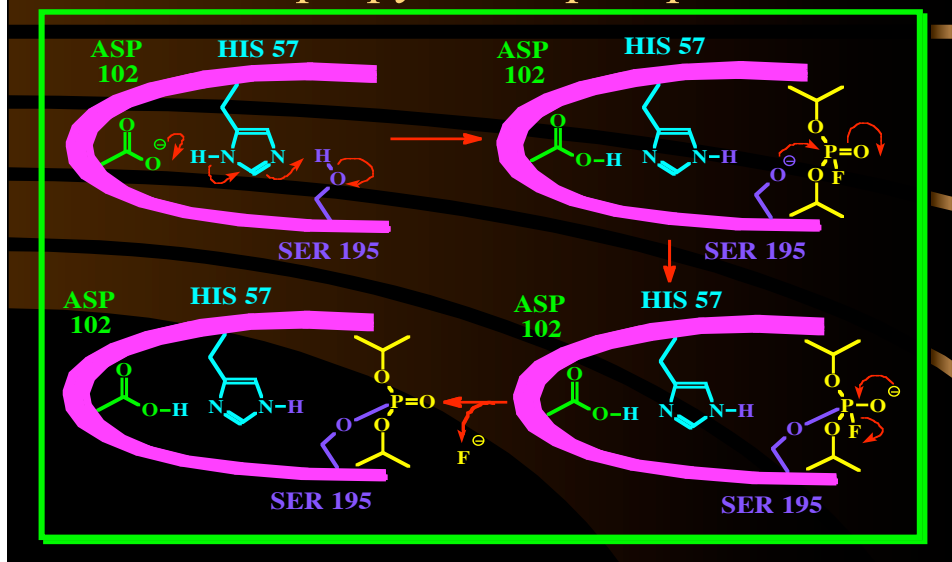
Blocking of lysine with TNBS



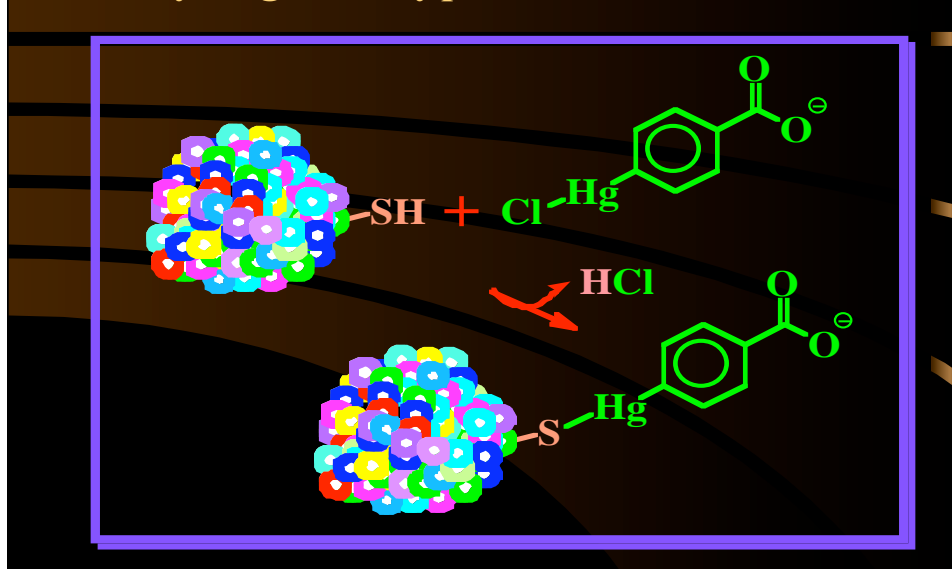
Modification of lysine



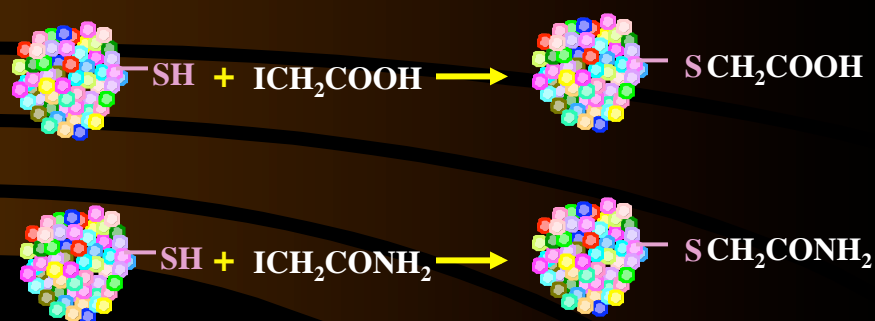
Chymotrypsin inactivation by diisopropylfluorophosphate



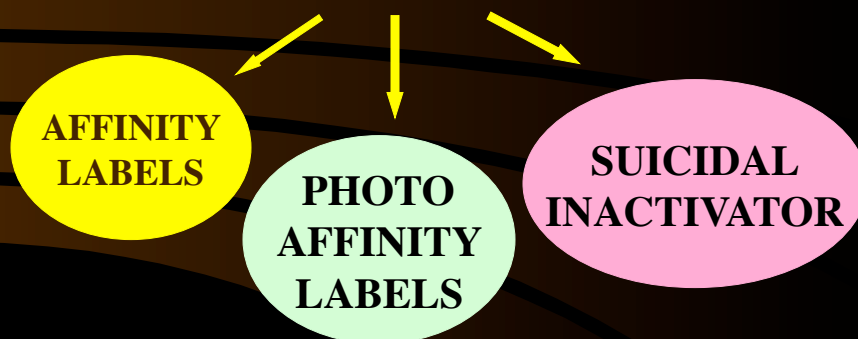
Inactivation of glyceraldehyde-3-phosphate dehydrogenase by *p*-chloromercuribenzoate



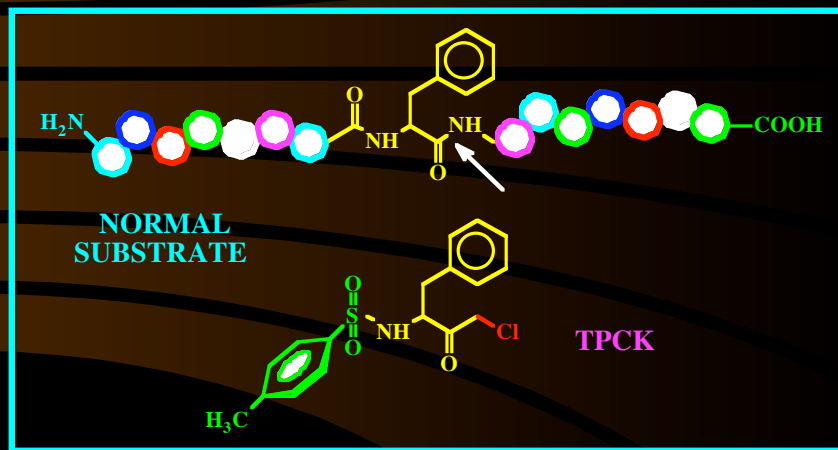
Iodoacetamide and Iodoacetic acid react with SH, imidazole and NH₂



SPECIFIC COVALENT INHIBITORS

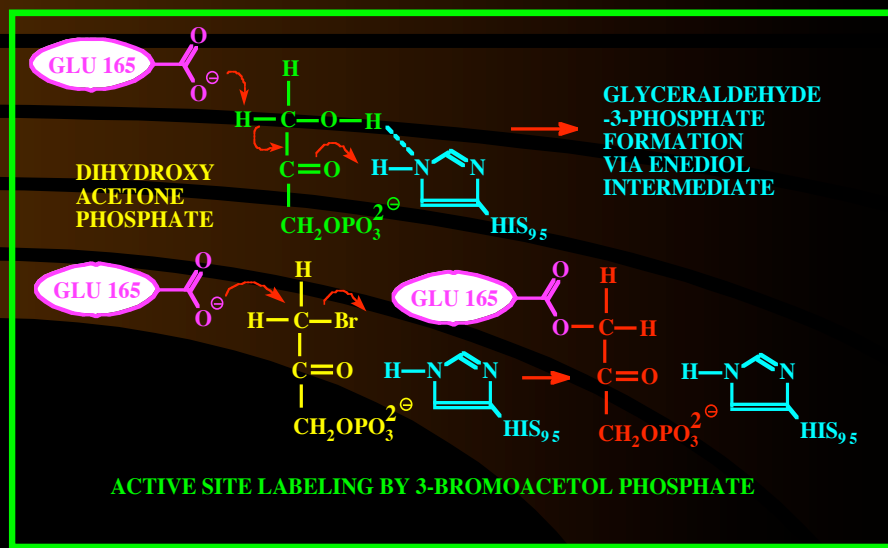


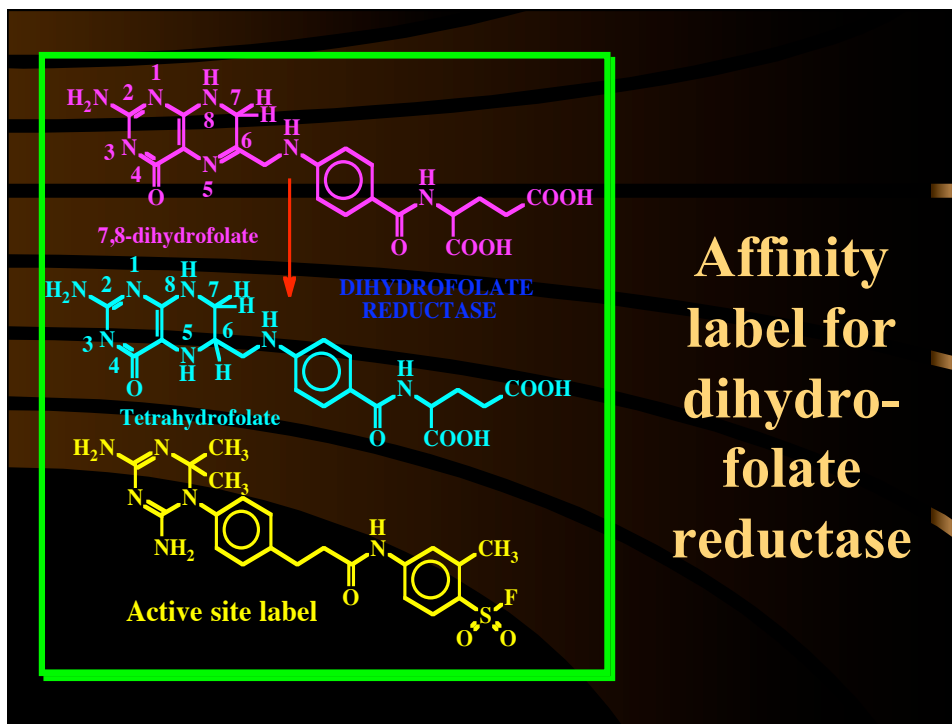
Affinity Label - TPCK



Top : Normal Chymotrypsin substrate
 Bottom : *p*-toluenesulfonylphenylalanylchloromethylketone
 Arrow indicates the site of cleavage

TRIOSE PHOSPHATE ISOMERASE REACTION -AFFINITY LABELLING



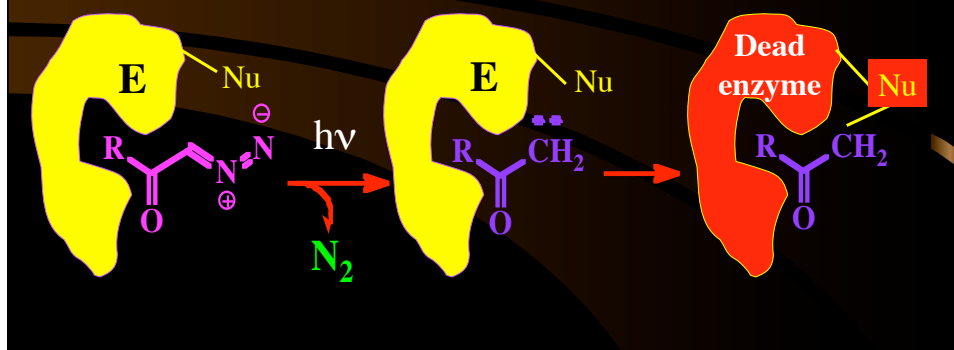


PHOTOAFFINITY LABEL

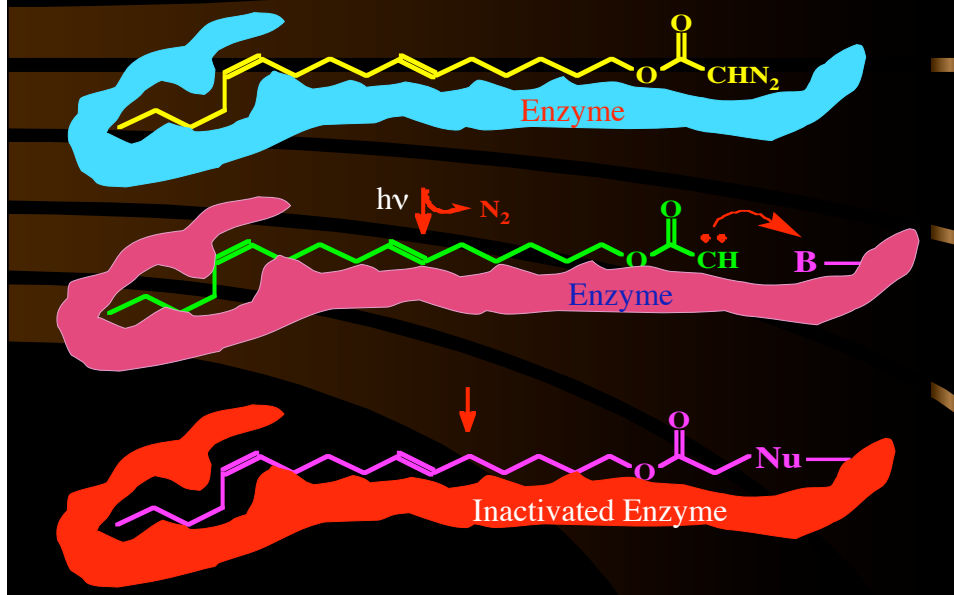
- Attach a photoreactive group such as diazoacetyl group to a substrate analog.
- Allow the enzyme bind to the photoaffinity label.
- Shine light on the complex.
- The photolabile group is activated and a reactive intermediate is generated.
- The reactive intermediate rapidly inactivates the enzyme by binding to essential amino acids near near the active site.

PHOTOAFFINITY LABEL - HOW IT WORKS

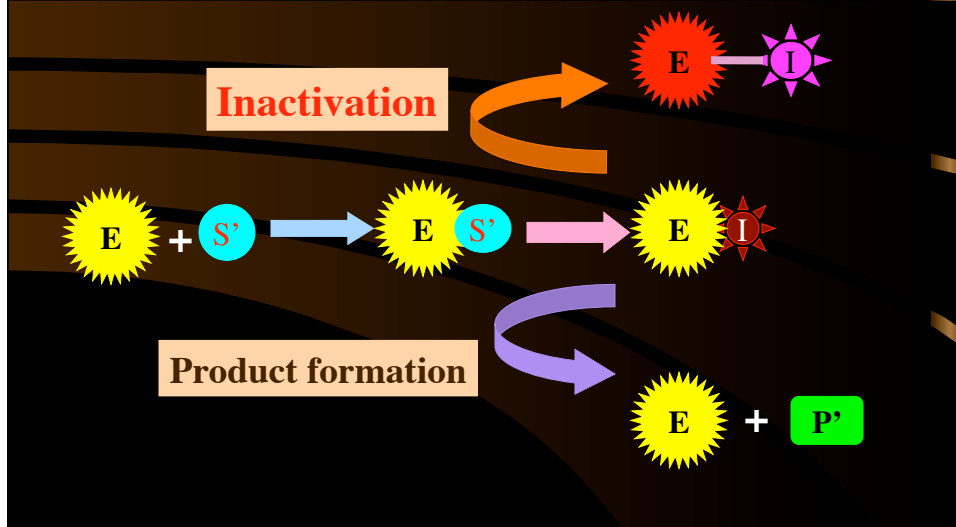
- Step 1. Photoaffinity label binds to the protein.
 at a site directed by the substituent group (R).
- Step 2. Light is shined on the complex to eliminate the photo labile group.
- Step 3. The reactive group generated attaches itself to the nearby nucleophile killing the protein.



6E, 11Z-hexadecadienyl diazoacetate interaction with pheromone binding protein



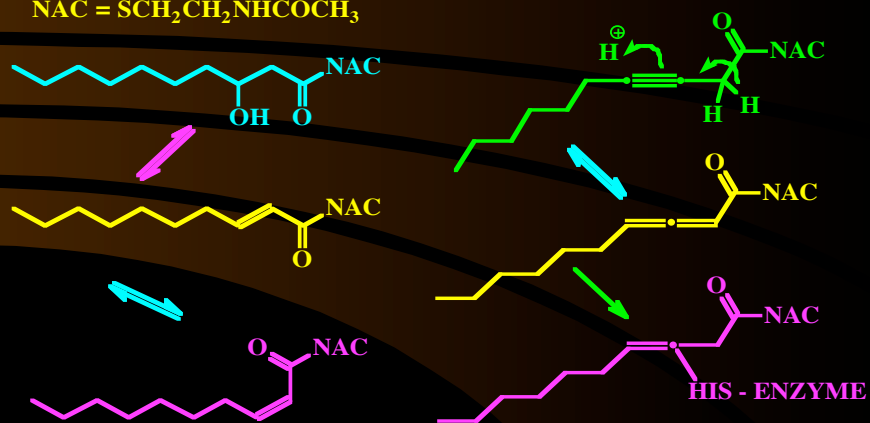
Mechanism of Suicidal Inactivation



First report on suicidal inactivation - Bloch's group

β -hydroxydecanoylthioester dehydrase

NAC = $\text{SCH}_2\text{CH}_2\text{NHCOCH}_3$



Criteria for suicidal inactivation

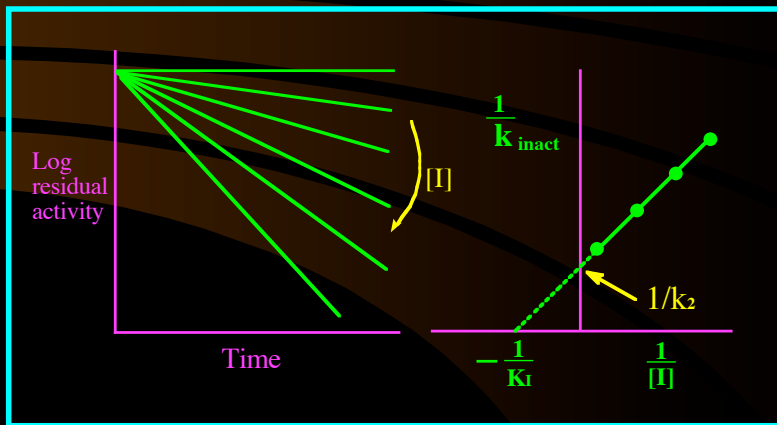
- **Inhibitor should be unreactive.**
- **It should become reactive only after activation by enzymatic action.**
- **Conversion of unreactive to reactive inhibitor should be due to specific catalytic function of the enzyme.**
- **Nonspecific interaction should be minimum.**
 - (a) Nontarget enzyme reaction
 - (b) Escape of reactive intermediate before reaction.

How to identify suicidal inactivators?

- **Best way to identify is the characterization of enzyme inhibitor complex by physico-chemical studies.**
- **But this could be difficult in some cases. So one needs some easy techniques to identify the suicidal inactivators.**

Kinetic experiments can identify suicidal inactivators

- The loss of enzyme activity should follow time dependent first order kinetics at fixed concentration of inhibitor.
- (If a reactive inhibitor comes out and then reacts with the enzyme, it will obviously follow second order kinetics. More over second addition of enzyme to this reaction mixture will cause an increased rate of inactivation due to accumulated inhibitor. Also addition of nucleophiles will reduce the rate of inactivation in this mode).



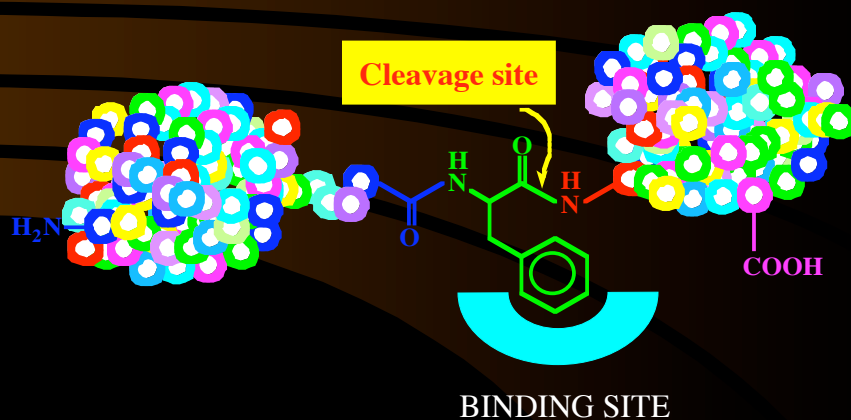
Kinetics of inactivation

- The rate of inactivation should follow Michaelis Menten type kinetics.
- Substrate (or competitive inhibitor) should protect the enzyme from inactivation.

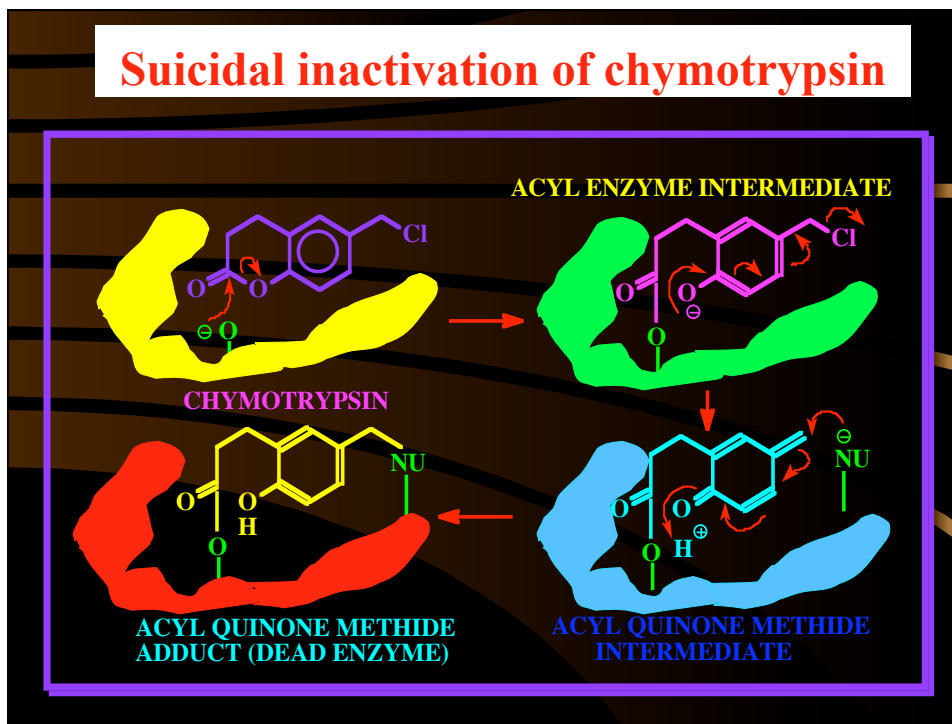
Inactivation Kinetics

- Inactivation Should be irreversible. (Show by dialysis, gel filtration etc.,)
- Inactivator should be bound covalently to the enzyme (show with radiolabel or other techniques).
- Stoichiometry of binding should be one to one mole.
- Partitioning between catalysis and inactivation should be determined.

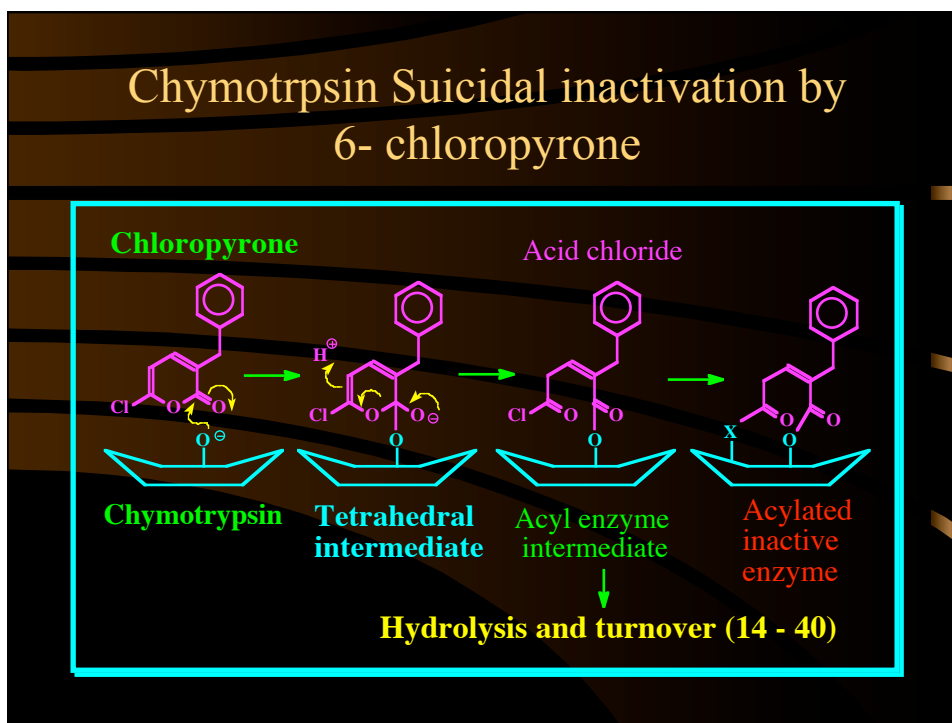
Chymotrypsin is an endopeptidase that cleaves proteins on the carboxyl side of aromatic amino acids (Phe, Tyr, Trp) and Leu.



Suicidal inactivation of chymotrypsin

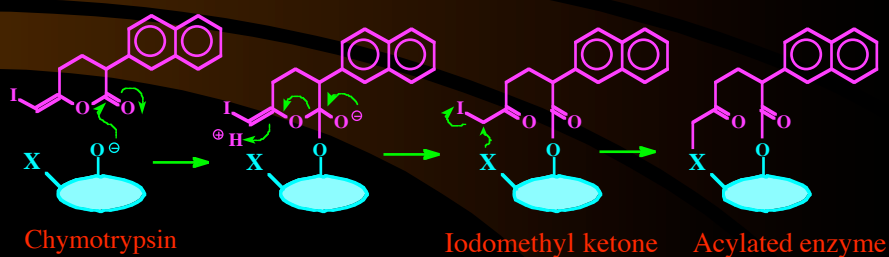


Chymotrpsin Suicidal inactivation by 6- chloropyrone



Highly specific chymotrypsin inactivator

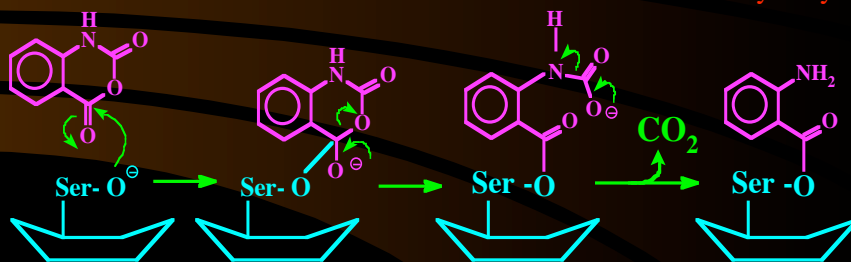
6-iodomethylene naphthyl tetrahydropyran-2-one (1.7 turnover per inactivation)



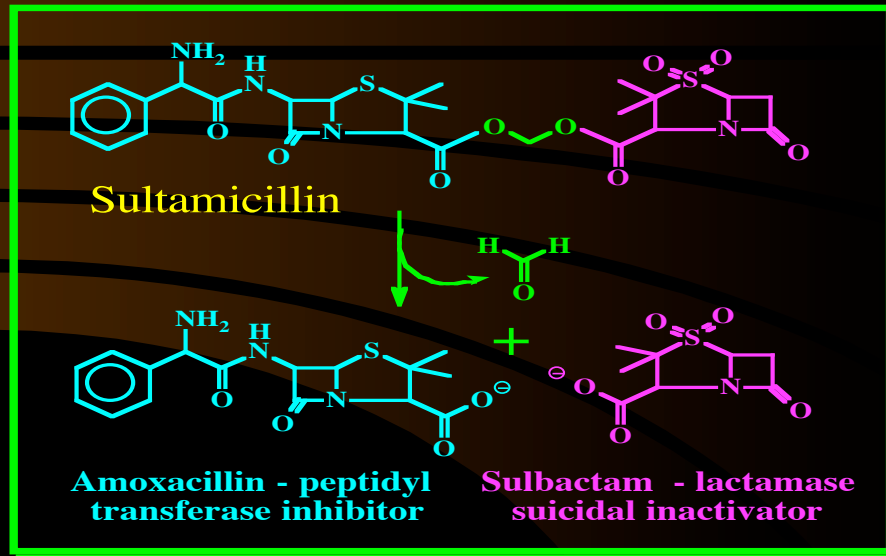
Stoichiometric titrant for chymotrypsin

Isatoic anhydride

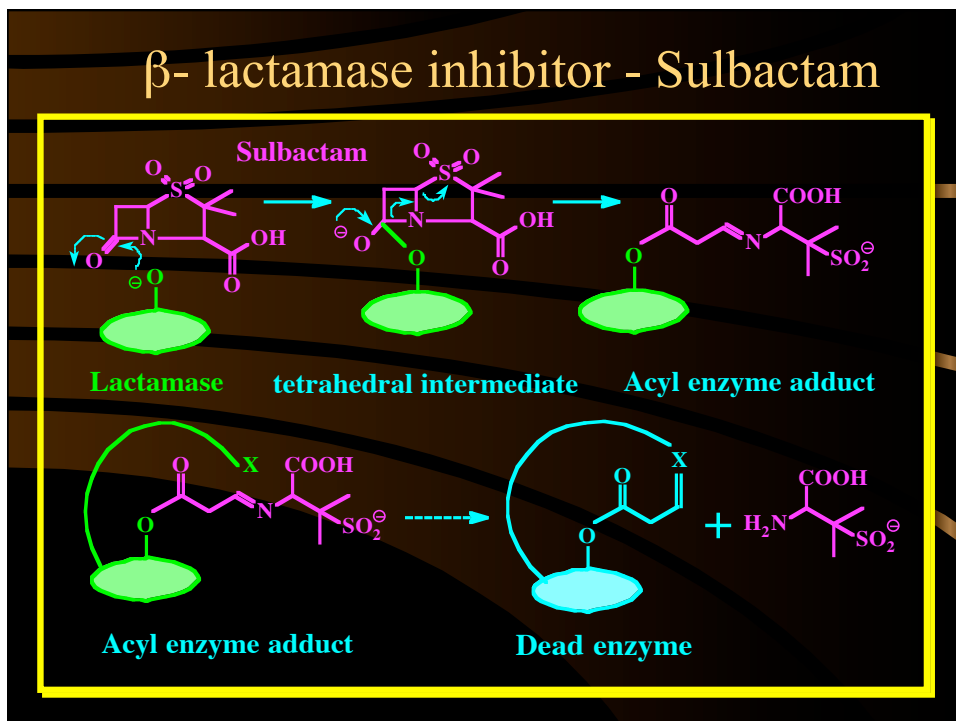
Anthranilyl enzyme
resistant to hydrolysis



β - lactamase inhibitor - Sultamicillin

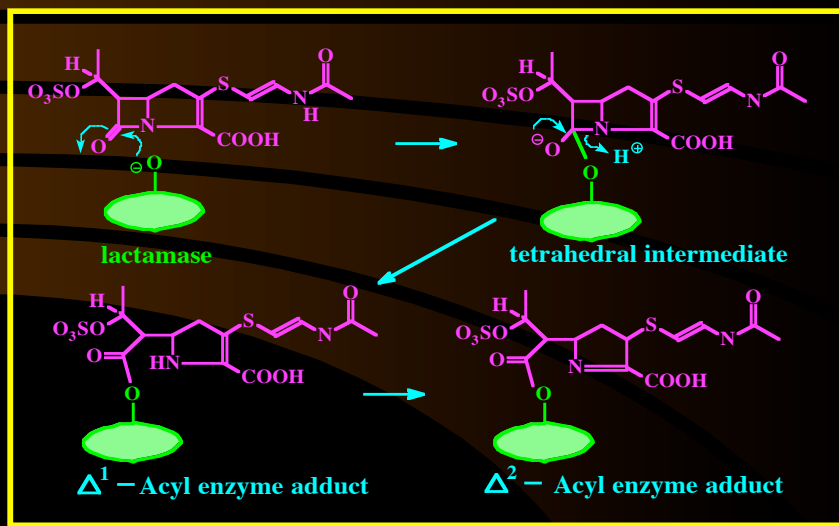


β - lactamase inhibitor - Sulbactam



β - lactamase inhibitor - Olivanate

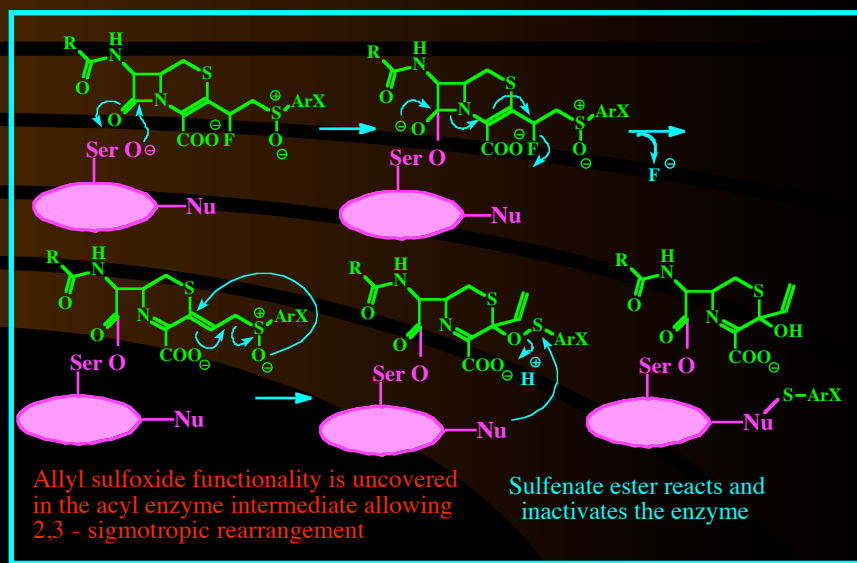
Both natural and synthetic carbapenems containing pyrroline ring inactivate lactamase by the following mechanism.



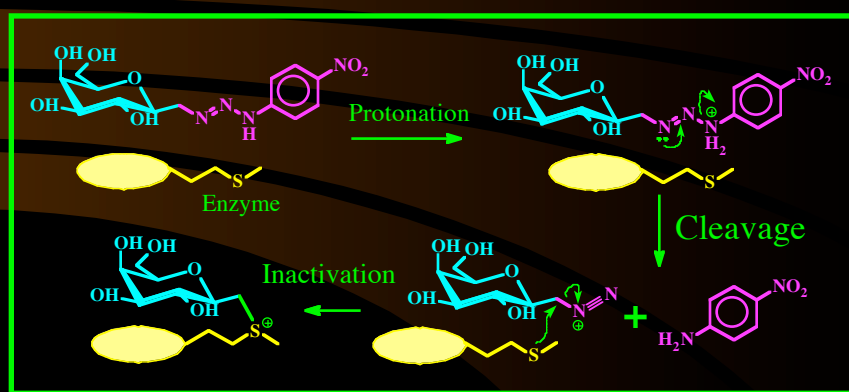
Lactamase suicidal substrate

One mole of acetyl methylene penicillanic acid kills lactamase with IC_{50} value of 1.4×10^{-9} M.

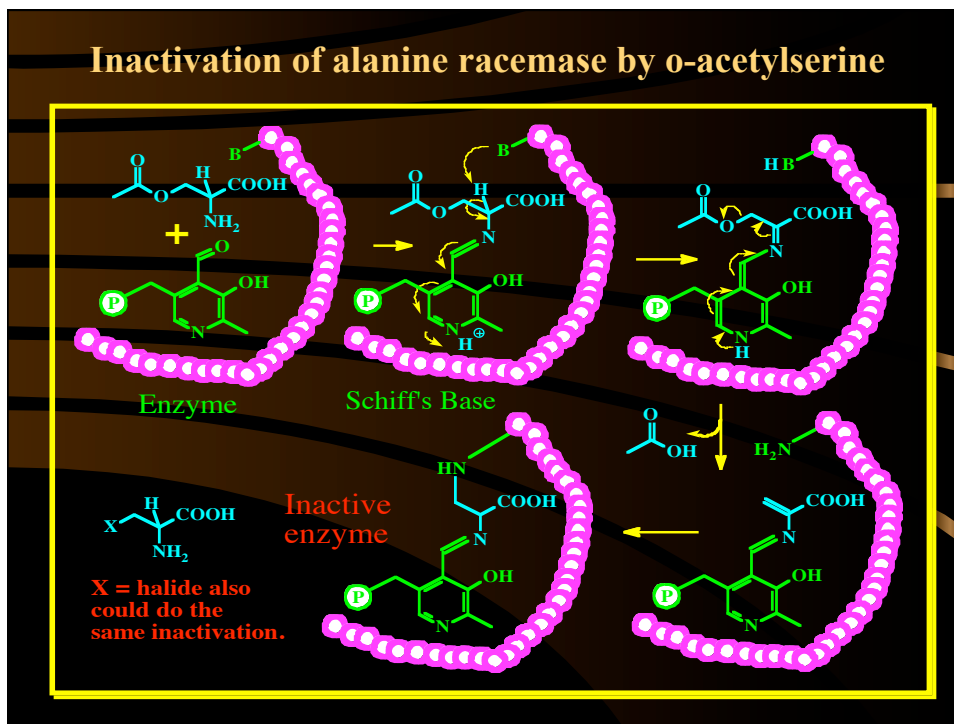
Cephalosporinase inhibitor based on 3'-exo substituent on cephalosporin hydrolysis



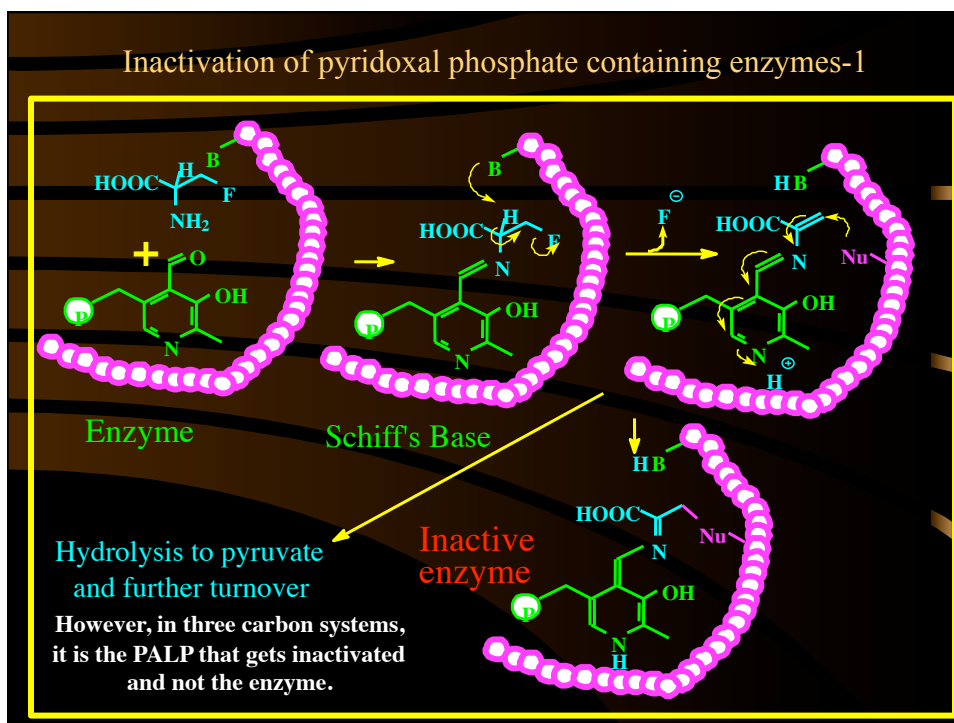
β -glycosidase inhibitor : β - D-galacto pyranosyl-*p*-nitrophenyltriazine in activated a β - glycosidase by adding on to the methionine at 500 position.



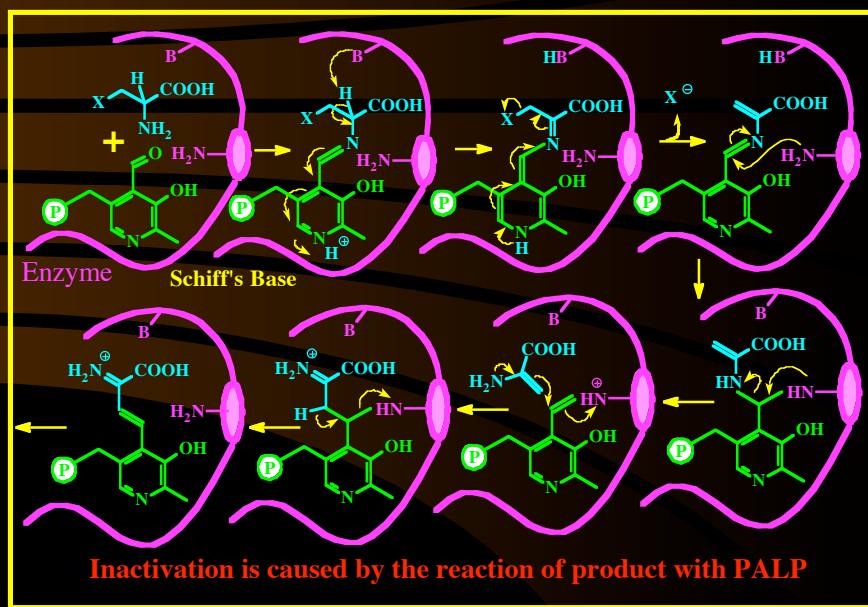
Inactivation of alanine racemase by o-acetylserine



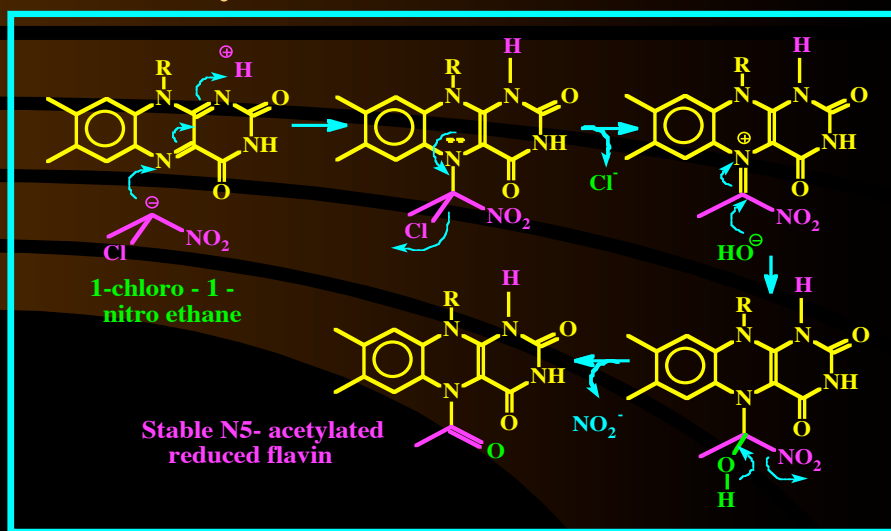
Inactivation of pyridoxal phosphate containing enzymes-1



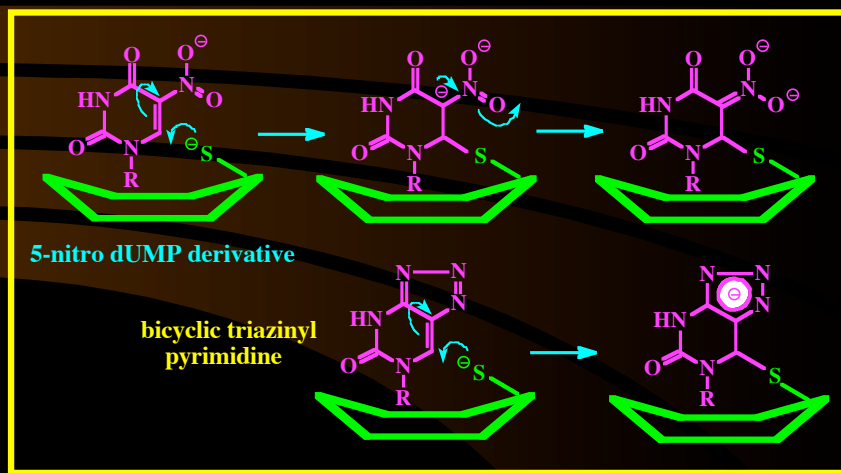
Formation of PALP adduct



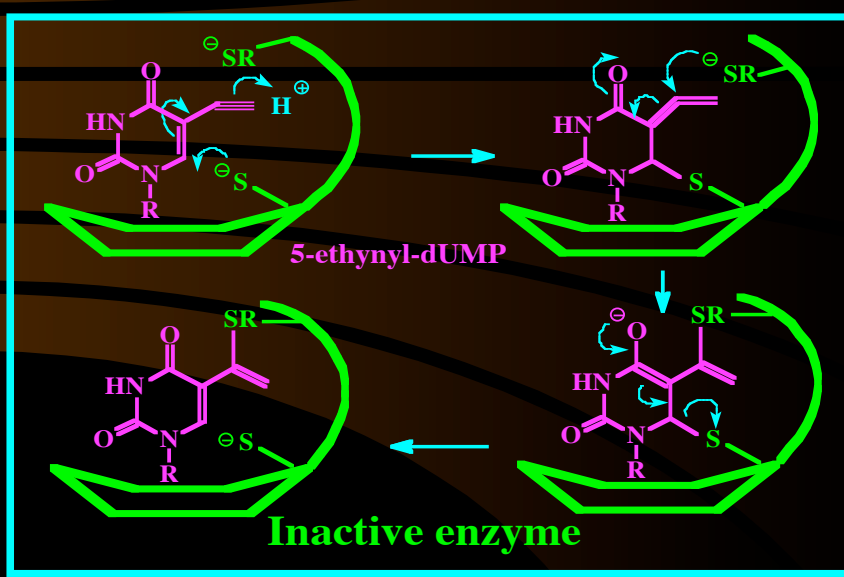
Inactivation of D-amino acid oxidase by 1-chloro-1-nitroethane



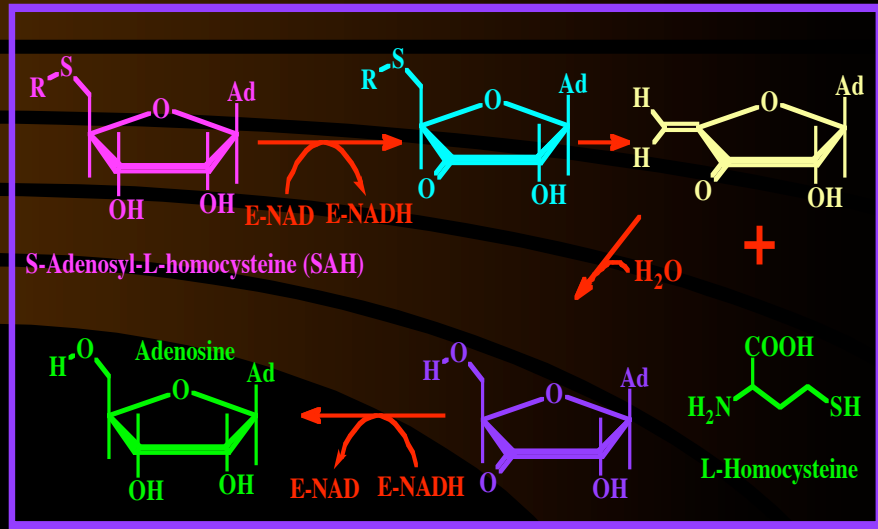
Thymidylate synthetase inactivators



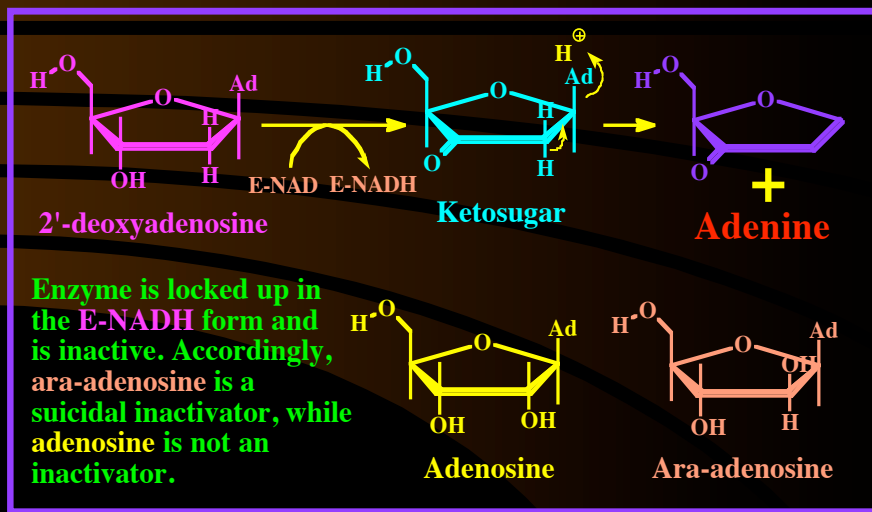
Thymidylate synthetase inactivator



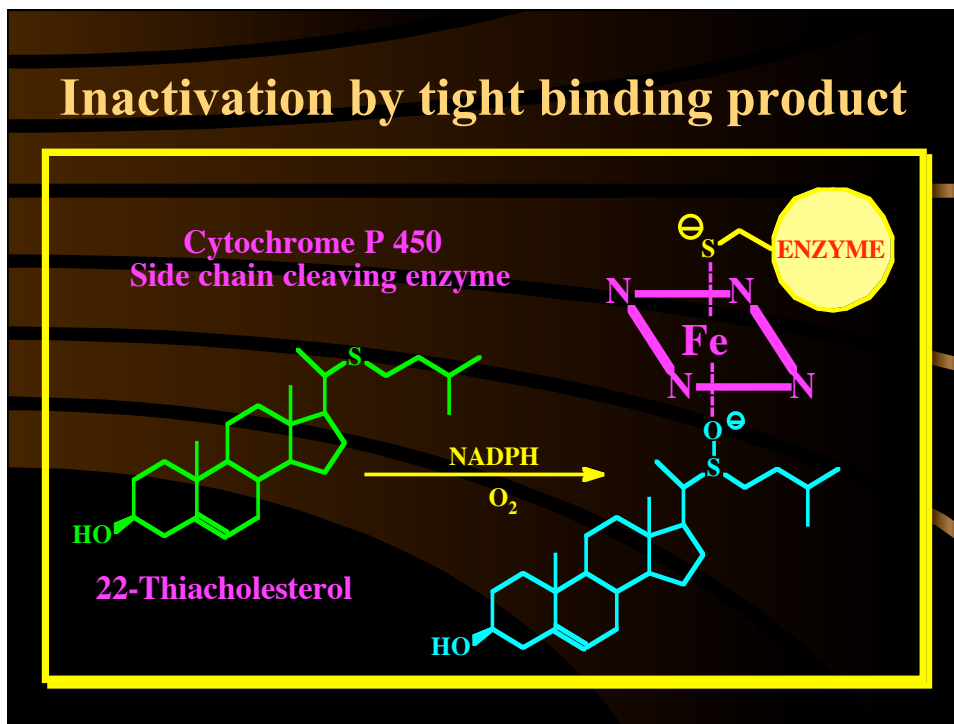
Reaction catalyzed by S-Adenosyl-L-homocysteinase



Inactivation of S-Adenosyl-L-homocysteinase by 2'-deoxyadenosine

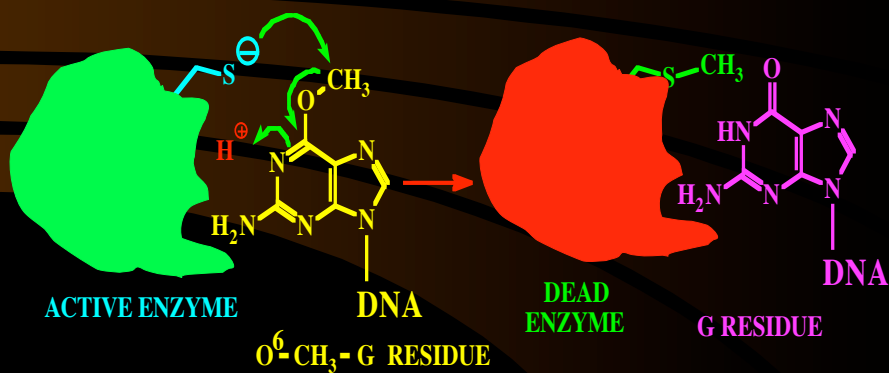


Inactivation by tight binding product



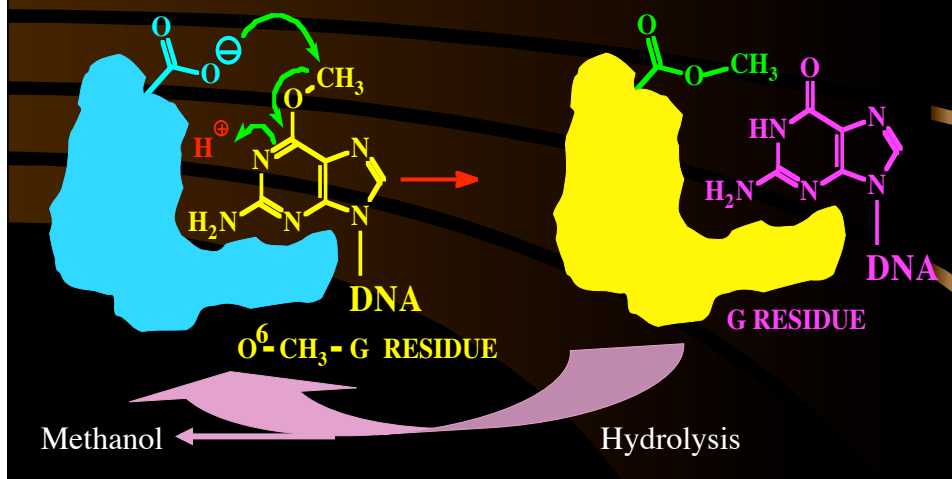
DEATH OF A REPAIR ENZYME

Enzyme uses a cysteine thiol group to abstract the methyl group.
The resultant thioether enzyme is irreversibly inactivated.

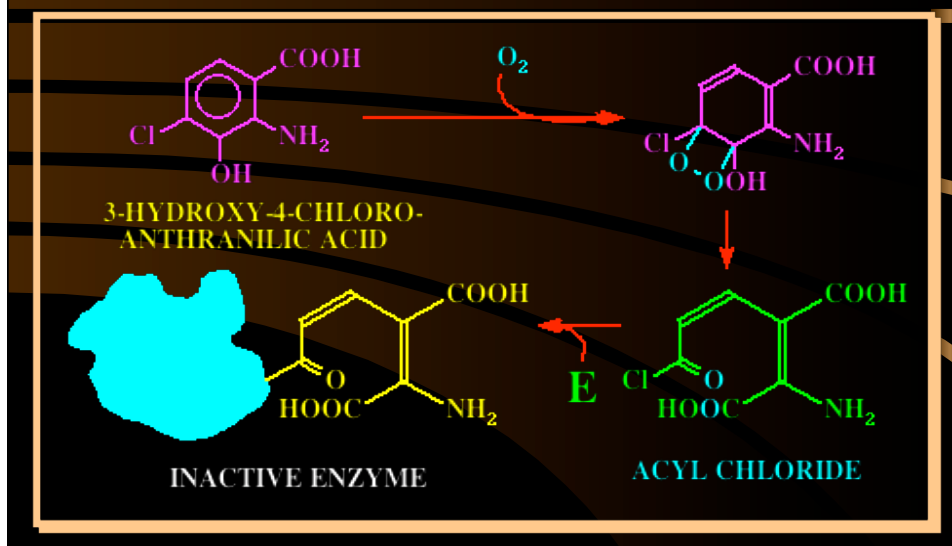


A SMART WAY TO MAKE THE ENZYME

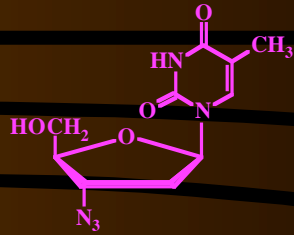
Instead of using cysteine as the active site amino acid, use aspartic acid (or glutamic acid). Once the carboxyl group abstracts the methyl group, use hydrolytic reaction to regenerate the enzyme.



Inactivation of 3-hydroxyanthranilate dioxygenase



AZT INHIBITS DNA POLYMERASE BY CHAIN TERMINATION MECHANISM



3'-azido-3'-deoxythymidine (AZT)

AZT gets converted into AZT triphosphate and is incorporated into the growing chain of DNA. But it lacks the 3' hydroxyl to allow further growth of the chain. Hence chain termination occurs.

